Gamma Irradiation of Human Platelet Lysate:

Validation of Efficacy for Pathogen Reduction and Assessment of Impacts on hPL Performance

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Introduction

Gamma irradiation is one of the most widely employed methods for pathogen reduction and commercial gamma sterilization facilities are easily accessible. The whole system for manufacturing gamma irradiated fetal bovine serum (FBS)¹ has been wellestablished, including dose range, dose mapping, frozen condition, as well as validation of pathogen reduction. Nevertheless, many research articles have addressed the optimal conditions for utilizing gamma irradiation in human plasma and blood components. With these comprehensive references, we previously assessed the feasibility of using gamma irradiation to obtain pathogen-reduced human platelet lysate (hPL) and reported low impacts on the potency for cell expansion. In this study, we validated the efficacy of gamma irradiation for virus inactivation. Four model viruses (BVDV, Reo3, HSV1, MMV) were chosen, per ICH/EMA guidelines^{2,3}, to represent a range of viruses with different genome, structure, size, and sensitivity to various chemical and physical agents. The virus spiked hPLs were gamma irradiated and the mean values of viral titers showed more than 4 log10 reduction across all model viruses. The results demonstrated gamma irradiation is an effective viral reduction procedure for hPL.

Results

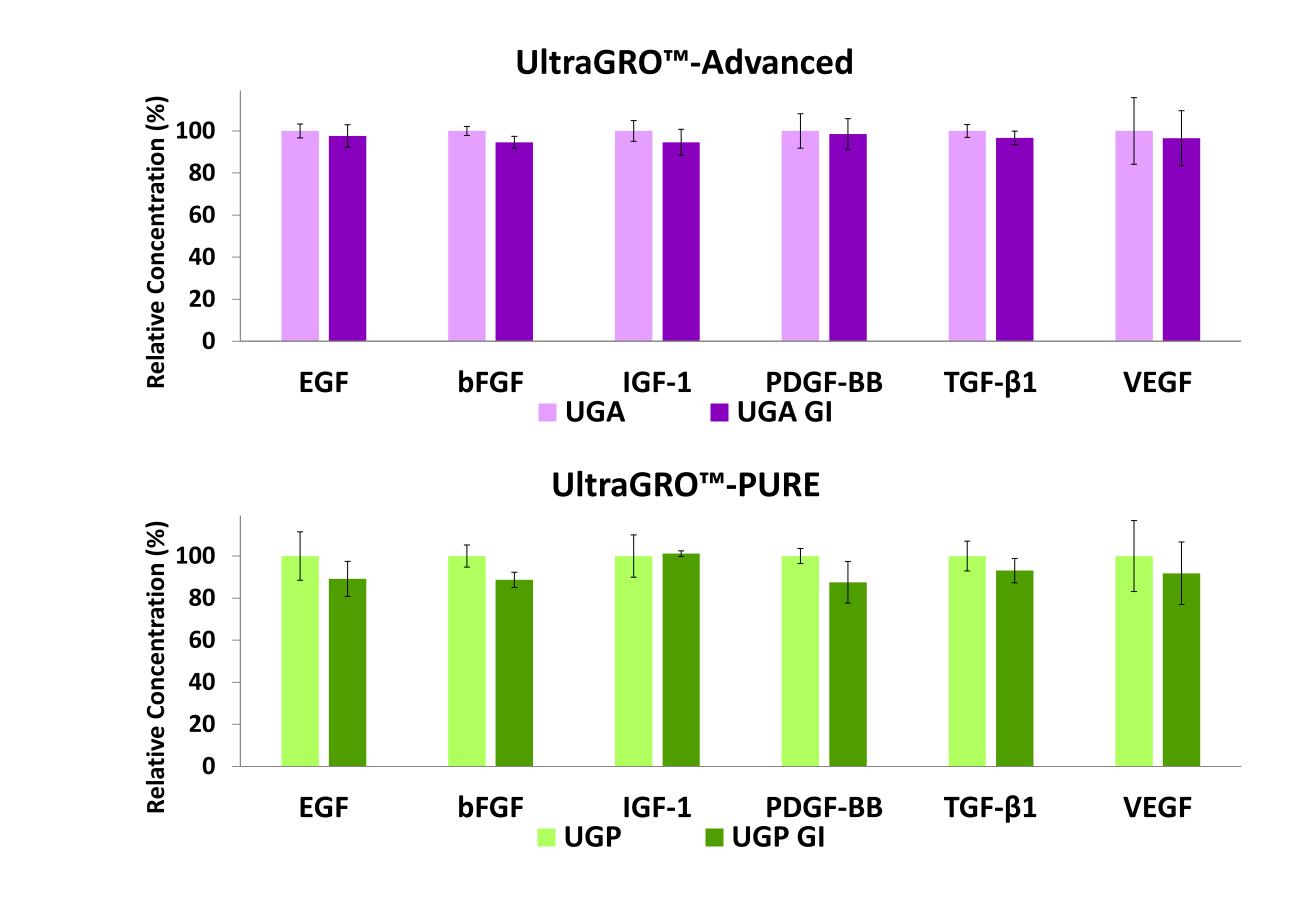
Gamma irradiation has low impact on cell expansion potency of UltraGRO[™]-Advanced & UltraGRO[™]-PURE

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ADMSC

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Growth factors retain comparable levels after gamma irradiation

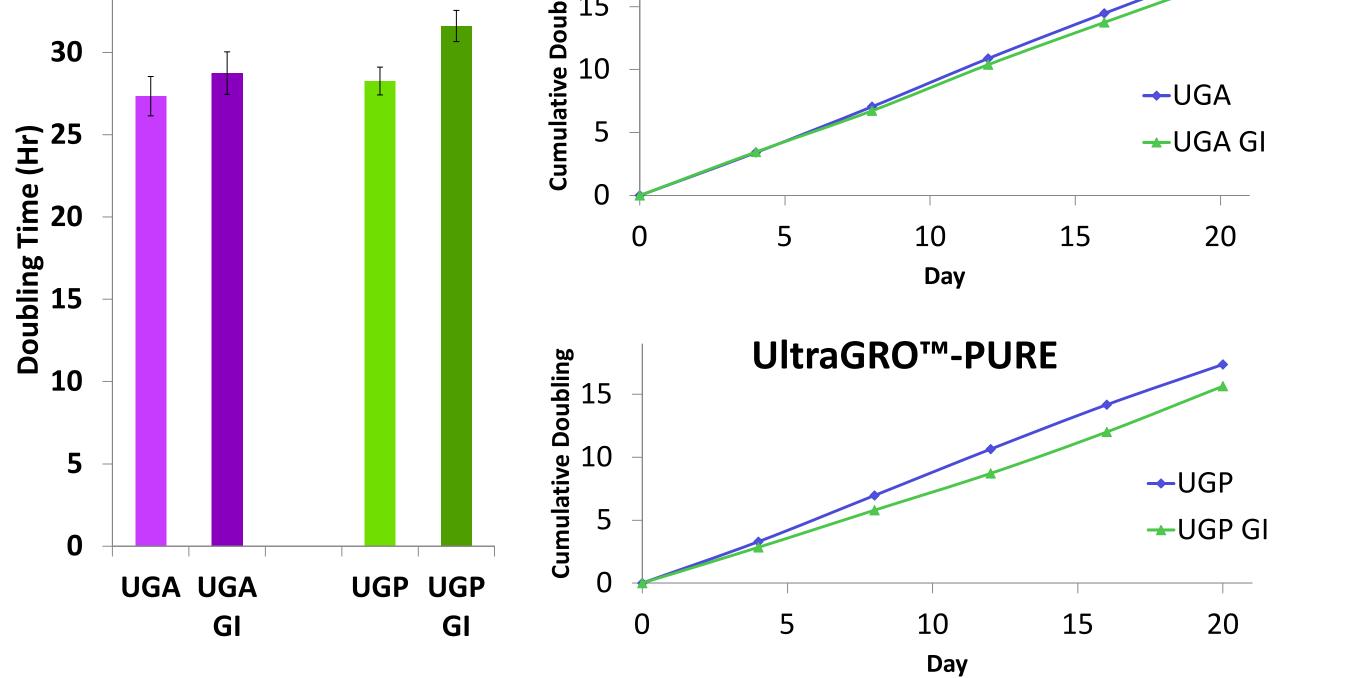


To assess the impacts of gamma irradiation on the long-term stability of hPL performance, we analyzed UltraGRO[™] GI series up to one year after gamma irradiation. The results showed growth factors still retained comparable levels to the non-irradiated hPLs. Mesenchymal stromal cells (MSC) cultured with gamma irradiated hPLs for more than three passages showed similar profiles as with the corresponding non-irradiated hPLs in respect of growth rate, morphology, immunophenotype, trilineage differentiation potency, and immunosuppressive property⁴.

Comparison of Pathogen Reduction Treatment (PRT)

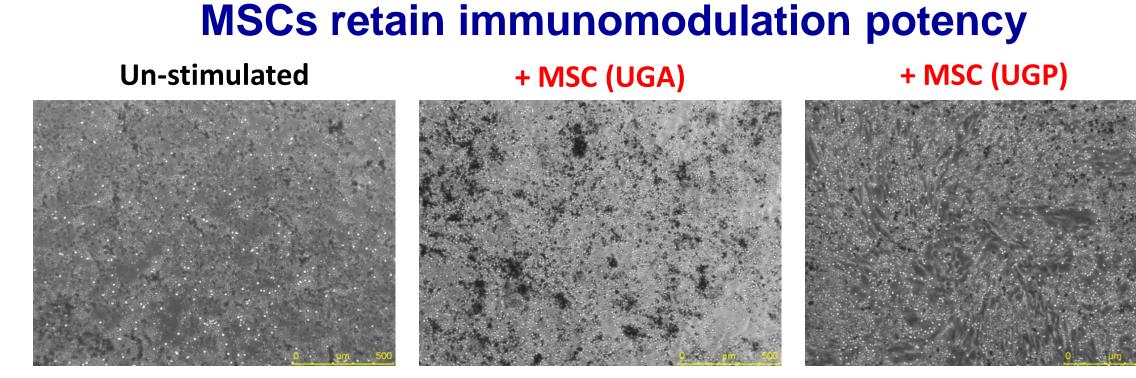
FBS vs UltraGRO™ GI series

| PRT FBS ¹ | VS | UltraGRO GI series | | |
|----------------------|--------------------|--------------------|--|--|
| 0.22µm | Sterile filtration | 0.22µm | | |



MSCs cultured with gamma irradiated supplements without significant change in immunophenotypes

| | Passage 5 | CD73 | CD90 | CD105 | CD34 | CD45 | CD11b | CD79a | HLA-DR |
|-----------|-----------|--------|--------------|-------|------|------|-------|-------|--------|
| UGA | ADMSC | 99.97 | 99.72 | 99.51 | 0.72 | 0.11 | 1.19 | 0.16 | 1.16 |
| | UCMSC | 99.03 | 99.90 | 99.94 | 0.36 | 0.03 | 1.85 | 0.34 | 1.89 |
| | BMMSC | 99.83 | 100.00 | 99.00 | 0.91 | 0.12 | 0.51 | 0.63 | 1.43 |
| UGA GI | ADMSC | 99.95 | 98.71 | 99.47 | 0.49 | 0.07 | 1.40 | 0.16 | 1.65 |
| | UCMSC | 98.12 | 99.94 | 99.91 | 0.25 | 0.12 | 1.94 | 0.51 | 1.97 |
| | BMMSC | 99.98 | 100.00 | 98.39 | 0.86 | 0.11 | 0.74 | 0.18 | 1.58 |
| UGP | ADMSC | 99.95 | 99.97 | 99.55 | 1.07 | 0.04 | 0.43 | 0.30 | 0.56 |
| | UCMSC | 99.08 | 99.91 | 99.91 | 0.11 | 0.98 | 1.46 | 0.31 | 0.74 |
| | BMMSC | 100.00 | 100.00 | 99.11 | 0.08 | 0.02 | 0.21 | 0.20 | 0.19 |
| UGP GI | ADMSC | 99.97 | 99.88 | 95.33 | 0.34 | 0.40 | 0.78 | 0.37 | 1.65 |
| | UCMSC | 95.51 | 99.98 | 99.09 | 0.80 | 0.31 | 1.08 | 1.11 | 1.97 |
| | BMMSC | 99.94 | 99.50 | 99.95 | 0.93 | 0.15 | 0.15 | 0.34 | 1.45 |



CD3/CD28 beads stimulation + MSC (UGA GI)

+ MSC (UGP GI)

| Finished products storage | -20 °C | |
|--|---|--|
| Transportation to irradiation plant | Frozen on dry ice | |
| Irradiation | Gamma | |
| Radiation source | Cobalt-60 | |
| Dosage | 25-40 kGy | |
| Physical state | Sealed bottles | |
| Temperature control | Dry ice | |
| Transportation to supplier storage | Frozen on dry ice | |
| | Transportation to irradiation plant Irradiation Radiation source Dosage Physical state Temperature control Transportation to | |

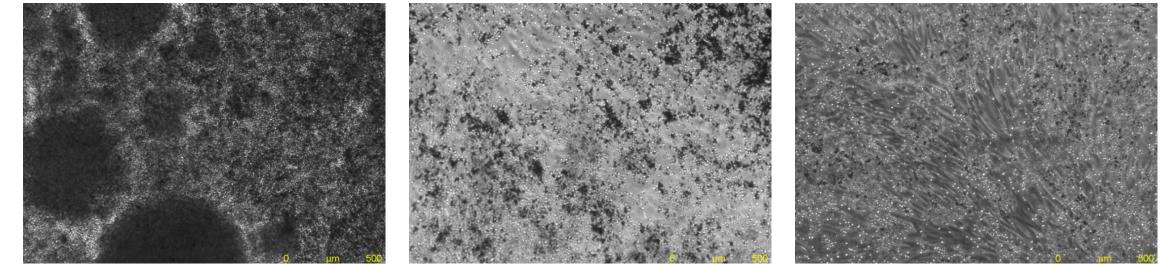
Viral Clearance Validation

| | RNA | RNA | DNA | DNA |
|--------------------------|-----------|-------------------|------------------|-------------------|
| Virus Category | Enveloped | Non- enveloped | Enveloped | Non- enveloped |
| Model for | HCV, HIV | HAV | CMV, EBV, HBV | B19 |
| Virus | BVDV | Reo3 | HSV1 | MMV |
| Family | Flavi | Reo | Herpes | Parvo |
| Genome | ssRNA | dsRNA | dsDNA | ssDNA |
| Size (nm) | 40-60 | 60-80 | 120-200 | 18-24 |
| Resistance | Low | Med-High | Medium | Very High |
| UltraGRO- PURE GI | > 5.42 | > 4.40 | > 4.51 | 4.55 |
| UltraGRO- Advanced GI | > 5.54 | > 4.27 | > 4.50 | 4.46 |

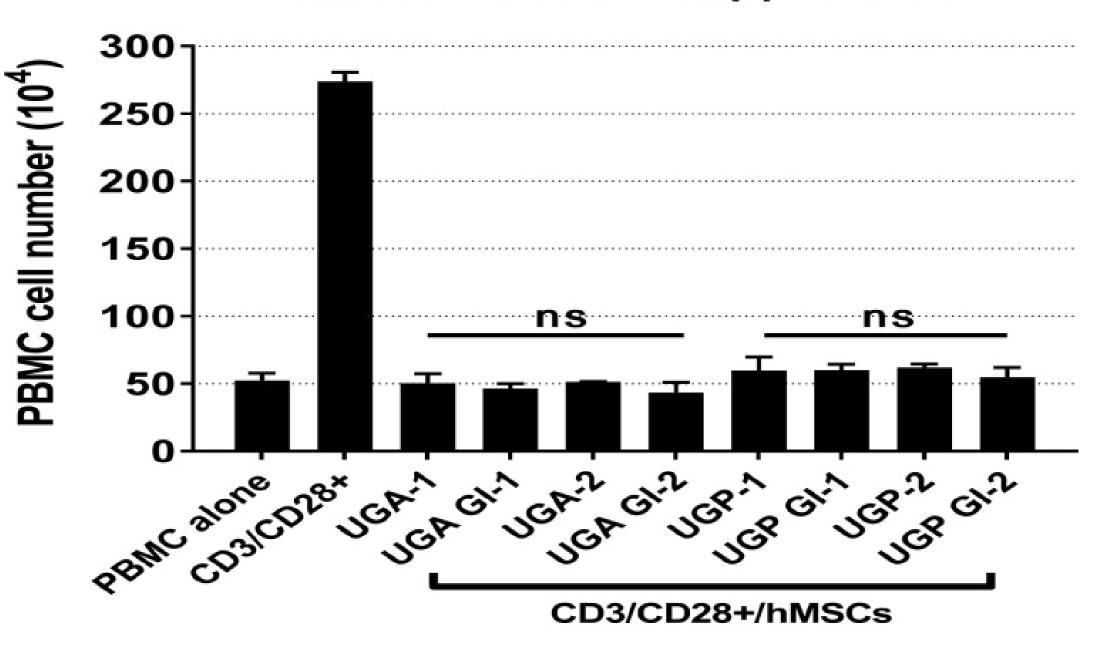
MSCs retain tri-lineage differentiation capability

ADMSCs P3
Adipogenesis
Osteogenesis
Chondrogenesis

UGA
Image: Construction of the second of the secon

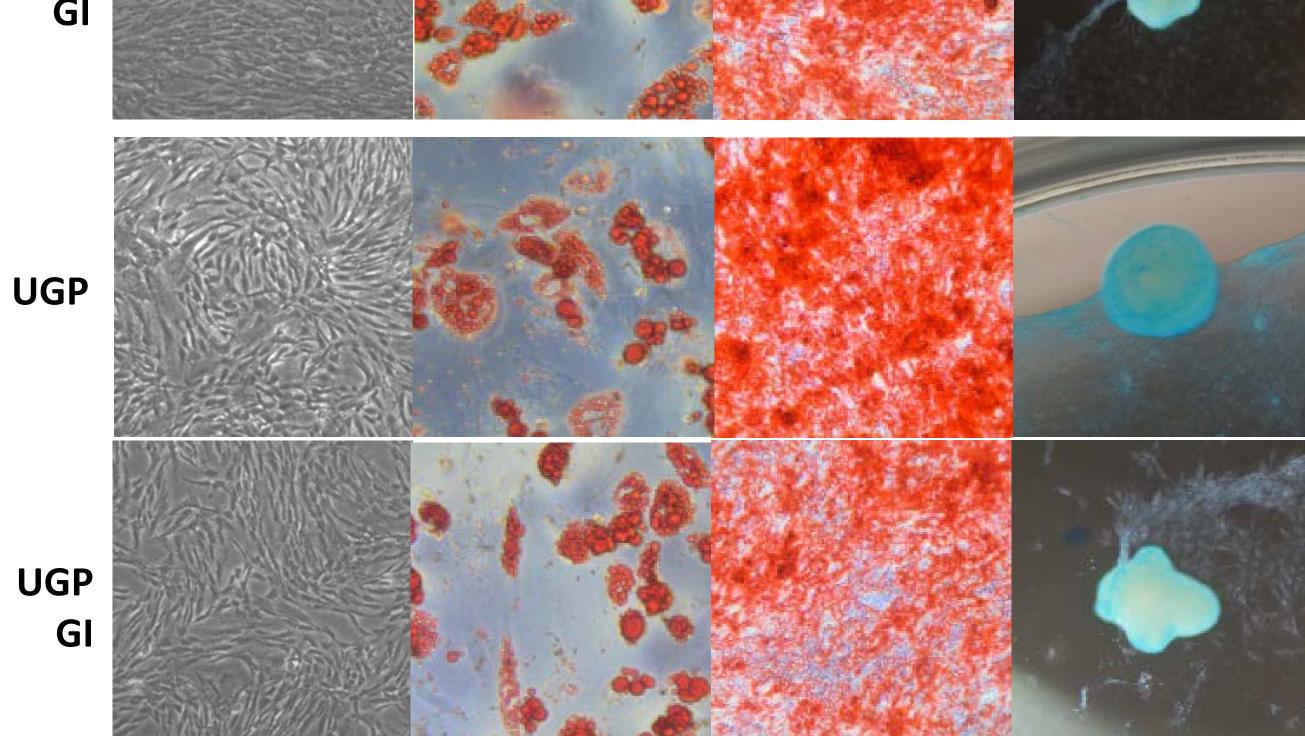


hMSC Immunosuppression



Conclusions

Gamma irradiation can be considered as a feasible approach for pathogen reduction treatment of pooled human platelet lysate. While



gamma irradiation make hPL to be a safer cell culture supplement, we demonstrate gamma irradiated UltraGRO[™] GI series products retain the potency to support cell growth and have low impact on key growth factors. UltraGRO[™] GI series is an ideal FBS substitute for cell therapy and regenerative medicine applications.

Reference

- 1. Gamma irradiation of animal sera for inactivation of viruses and mollicutes Review. Biologicals 2011, 39: 370-377.
- 2. Viral Safety Evaluation of Biotechnology Products Derived from Cell Lines of Human or Animal Origin Q5A (R1)" (ICH, 1999)
- 3. Revised "Note for Guidance on Virus Validation Studies: The Design, Contribution and Interpretation of Studies Validating the Inactivation and Removal of Viruses" (CPMP/BWP/268/95, European Medicines Agency, 1996)
- 4. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. Cytotherapy 2006, 4: 315–317.